Contribution from the Department of Chemistry and Biochemistry, Utah State University, Logan, Utah 84322

Reduction of Nitrate by Diammonium Oxopentachloromolybdate(V) in Dimethylformamide

R. D. TAYLOR and J. T. SPENCE*

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The reduction of nitrate by diammonium oxopentachloromolybdate(V) ((NH4)₂MoOCl₅) in anhydrous dimethylformamide (DMF) has been investigated as a model for biological nitrate reduction. The stiochiometric reaction is MoOCl₅²⁻ + NO₃⁻ \rightarrow MoO₂Cl₂ + NO₂ + 3Cl⁻. The reaction is first order in total Mo(V) and NO₃⁻ and is inhibited by Cl⁻. A mechanism involving the formation of an intermediate complex, MoOCl₃NO₃⁻, followed by an intramolecular electron transfer to form products has been developed to explain the kinetic results. A rate constant of (1.62 ± 0.15) × 10⁻⁴ sec⁻¹ for the overall reaction has been obtained and activation parameters of 18.3 ± 0.4 kcal mol⁻¹ for ΔH^{*} and -14 ± 3 cal mol⁻¹ K⁻¹ at 25° for ΔS^{*} determined. The implications for enzymatic nitrate reduction are discussed.

The major source of nitrogen for most plants and microorganisms is nitrate, which is reduced in a number of steps by the organisms to ammonia. The first stage in this process, the reduction of nitrate to nitrite, is catalyzed by a group of enzymes, the nitrate reductases, all of which require molybdenum as a cofactor.¹ In addition to molybdenum, flavine,² *b*-type cytochromes,² and nonheme iron cofactors^{3,4} may also be required, depending on the specific enzyme.

Little is known concerning the binding site of molybdenum, its oxidation states, or its function in the catalytic processes of the nitrate reductases. Early work suggested the metal shuttles between the +5 and +6 oxidation states, transferring electrons from a reduced flavine cofactor to nitrate.⁵ More recently, electron spin resonance (ESR) studies of the enzymes from *M. denitrificans*³ and *E. coli*⁴ have shown the presence of Mo(V) during the catalytic cycle. Evidence from both enzymatic and model investigations suggests the protein at the molybdenum binding site provides a nonaqueous environment similar to an aprotic medium.^{1,6,7}

Previous attempts to reduce nitrate by molybdenum(V) species in aqueous solution have produced mainly nitric oxide,^{8,9} due to the more rapid reduction of nitrite than nitrate by Mo(V) in water.^{8,10} A recent preliminary report indicates NO₂ is formed upon reduction of nitrate by molybdenum(V) complexes in dichloromethane solution.¹¹

If the environment at the molybdenum binding site is indeed nonaqueous, an aprotic solvent of moderate dielectric constant should provide a more appropriate environment than water for model studies of nitrate reduction. As part of a program of model studies for molybdenum enzymes, the reduction of nitrate by diammonium oxopentachloromolybdate(V), (NH4)2MoOCl5, in anhydrous dimethylformamide (DMF) is reported here.

Experimental Section

Materials. Spectrograde DMF was obtained from Aldrich Chemical Co. It was distilled in vacuo over P₂O₅ (2.5–8.0 Torr, $33-49^{\circ}$) and stored over Linde molecular sieves, Type AW-500, for at least 48 hr before use. This technique is reported to reduce the H₂O content to <18 ppm.¹²

(NH4)2MoOCl5 was prepared and purified as described.¹³ Tetraethylammonium nitrate (Et4NNO3) and NaNO3, Eastman Kodak and Mallinckrodt products, respectively, were dried in vacuo over P2O5 before use. MoO2Cl2 was obtained from Alfa Products and used without further purification. NO2 and N2 (99.99%) were obtained from Matheson. NH4MoOCl4-CH3CN was synthesized according to the method of Horner and Tyree.¹⁴

Methods. Stock solutions of $(NH_4)_2M_0OCl_5$ and Et4NNO3 or NaNO3 were prepared in deaerated DMF and kept under a stream of N₂ in a constant-temperature bath. The reaction was initiated by mixing the proper amounts of stock solutions, using gas-tight syringes, under N₂ in a cuvette equipped with a rubber septum. The cuvette

was kept in a thermostated cell holder in a Varian 635 spectrometer and the absorbance of (NH4)₂MoOCl₅ at 315 nm as a function of time was recorded on a Hewlett-Packard X-Y recorder. The method was standardized by preparing known solutions of (NH4)₂MoOCl₅ in DMF and measuring their absorbance at 315 nm. Over the concentration range used $(10^{-5}-10^{-4} M)$ Beer's law is obeyed (ϵ 5.36 × 10³). Absorbance by NO₃⁻ is negligible at 315 nm while small corrections for the absorbance of NO₂ and MoO₂Cl₂ were necessary (ϵ NO₂ + ϵ MoO₂Cl₂ = 633) at the concentrations used or expected to be formed in the reaction. The molar absorptivity (5.23 × 10³) at 315 nm of solutions of (NH4)₂MoOCl₅ in DMF was unaffected by added Et₄NCl concentrations up to 2.5 × 10⁻² M.

Except in cases where NO₂ was to be detected or HNO₂ measured, N₂ was continuously passed over the solution in the cuvette during the reaction. For HNO₂ analysis, samples were withdrawn with a small gas-tight syringe and added to an aqueous acidic solution for HNO₂ determination by the method of Shinn.¹⁵ Under these conditions, the NO₂ rapidly disproportionates to NO₃⁻ and HNO₂.^{8,16} Product identification was accomplished by preparing solutions of MOO₂Cl₂ and NO₂ and comparing their absorption spectra with the spectrum of the reaction mixture.

In all cases, extreme care was necessary to prevent the absorption of H_2O and O_2 by the solutions, since these react with (NH_4)₂MoOCl₅, giving unreproducible results.

All kinetic data were treated with appropriate programs on a PDP-8 computer to give rate constants and activation parameters.

Results

Stoichiometry. When nitrate (tetraethylammonium nitrate, Et4NNO3, or NaNO3) is added to a solution of $(NH4)_2$ -MoOCl5 in DMF, the characteristic spectrum of $(NH4)_2$ -MoOCl5 changes with time to that of a solution of MoO₂Cl₂ and NO₂. Samples, withdrawn from the reaction, were tested for nitrite in acidic aqueous solution, while the amount of $(NH4)_2MoOCl_5$ reacted was determined from spectrophotometric analysis of the original solution. The ratio of $(NH4)_2MoOCl_5$ reacted to HNO₂ formed was found to be 2.2 \pm 0.2. Furthermore, when the rate of the reaction was determined by following HNO₂ production or $(NH4)_2MoOCl_5$ disappearance, the results were essentially identical (see below). Thus, the stoichiometry as determined by analysis in acidic aqueous solution is

 $2M_0OCl_3^2 + NO_3 + 5H_2O \rightarrow HNO_2 + 2H_2M_0O_4 + 10C\Gamma + 5H^+$

(Mo(VI) is indicated as H2MoO4; in acidic solution, however, it is polymerized to some extent.)

The $\dot{H}NO_2$ arises by the well-known disproportionation of NO₂ in H₂O^{8,16}

 $H_2O + 2NO_2$ (or N_2O_4) $\neq NO_3^- + HNO_2 + H^+$

This was verified by bubbling NO_2 through DMF and testing the solution for HNO_2 in the same manner.

The stoichiometric equation in DMF may therefore be written

 $MoOCl_5^{2-} + NO_3^{-} \rightarrow MoO_2Cl_2 + NO_2 + 3Cl^{-}$

Kinetics. The rate of the reaction was determined by measuring the $(NH_4)_2M_0OCl_5$ concentration or the HNO₂ produced with time. The rates obtained were identical within experimental error.

The rate of the reaction was found to be dependent on both NO₃⁻ and (NH4)₂MoOCl₅ concentrations. Pseudo-first-order plots (excess NO₃⁻, first order in (NH4)₂MoOCl₅) were linear to approximately 50% reaction, deviating in a negative direction beyond this. Since negative deviations in kinetic plots are often indicative of product inhibition, additional chloride (Et4NCl) was added to the reaction, resulting in a decreased rate, while addition of MoO₂Cl₂ or NO₂ had no effect on the rate. Furthermore, as the additional chloride ion concentration was increased, the linearity of the plots increased beyond 50% and good pseudo-first-order plots were obtained when excess Et4NCl (>10⁻² M) was added.

Second-order rate constants, evaluated from the linear portion of the plots, gave consistent values when Et4NNO₃ was used as the source of nitrate but gave values that decreased with increasing nitrate concentration when NaNO₃ was used. This appears to be due to ion-pair formation between Na⁺ and NO₃⁻ in DMF, for which the ion-pair formation constant is reported to be¹⁷

$$\frac{[\text{Na}^+\text{NO}_3^-]}{[\text{Na}^+][\text{NO}_3^-]} = K_p = 43$$

When [NO₃⁻] calculated from this equilibrium is used in the rate expression, the rate constants are consistent and in good agreement with the rate constants for the results with Et₄NNO₃. Apparently, Et₄NNO₃ does not form ion pairs in DMF at the concentrations used. A search of the literature failed to find any data indicating pair formation for this compound.

Discussion

The best fit for the experimental data in the absence of added Cl⁻ was obtained by assuming an initial Cl⁻ concentration equivalent to the initial (NH₄)₂MoOCl₅ concentration. With this assumption, a rate expression was obtained (see below) that gives linear plots to >90% reaction.

The results are consistent with the mechanism of eq 1-3.

$$MoOCl_{5}^{2-} \xrightarrow{k_{1}}_{k_{-1}} MoOCl_{4}^{-} + Cl^{-} \qquad K_{1} = k_{1}/k_{-1}$$
(1)

$$MoOCl_4 + NO_3 \xrightarrow{k_1} MoOCl_3 NO_3 + Cl^-$$
(2)

$$MoOCl_3NO_3^{-} \xrightarrow{k_3} MoO_2Cl_2 + Cl^{-}$$
 (3)

Reaction 1 is a rapid equilibrium. Reaction 2 is the displacement of a second Cl⁻ by NO_3^- to form the MoOCl₃ $NO_3^$ complex. An intramolecular electron transfer of MoOCl₃ $NO_3^$ occurs in reaction 3 to produce the products. Application of the steady-state condition to MoOCl₃ NO_3^- leads to the rate expression

$$d[P]/dt = \frac{k_2 k_3 [MoOCl_4][NO_3]}{k_3 + k_2 [Cl]}$$

where $P = \text{products} = MoO_2Cl_2 = NO_2$. Assuming the absorbance at 315 nm measures the total Mo(V) concentration $(MoOCl_4^- + MoOCl_5^{2-})$ leads to the rate expression

$$d[P]/dt = -d[Mo(V)]/dt = \frac{k_2 k_3 K_1 [NO_3^{-}][Mo(V)]}{(k_3 + k_{-2} [Cl^{-}])(K_1 + [Cl^{-}])}$$



Figure 1. Plot of $[NO_3^-]/k''$ obsd [Cl⁻] vs. [Cl⁻] for runs with excess added Cl⁻ (25°).

If $k_3 \ll k_{-2}$ [Cl⁻], this reduces to

$$-d[Mo(V)]/dt = \frac{k_2 k_3 K_1 [NO_3^-][Mo(V)]}{k_{-2} [C\Gamma^-](K_1 + [C\Gamma^-])} = \frac{k_3 K_1 K_2 [NO_3^-][Mo(V)]}{[C\Gamma^-](K_1 + [C\Gamma^-])}$$

$$K_2 = k_2/k_{-2}$$

In the runs without added Cl⁻, it is assumed that $K_1 \gg [Cl^-]$, which reduces the expression to a form which can be integrated

$$-d[Mo(V)]/dt = \frac{k_3K_2[NO_3^-][Mo(V)]}{[C\Gamma]}$$

If $K_1 \gg [Cl^-]$, the initial [Cl⁻] \simeq initial [(NH4)₂MoOCl₅], and the integrated expression in the presence of excess NO₃⁻⁻ is

$$2[Mo(V)] - 3[Mo(V)]_0 \ln [Mo(V)] = k_3 K_2 [NO_3]t + I$$

where $[Mo(V)]_0$ = initial $[(NH_4)_2MoOCl_5]$, *I* is the integration constant, and $k_{obsd} = k_3K_2[NO_3^-]$. For the runs with NO₃⁻ not in excess (second order), the integrated expression is

$$2 \ln (a + [Mo(V)]) + \frac{3 [Mo(V)]_0}{a} \ln \frac{a + [Mo(V)]}{[Mo(V)]} = k_3 K_2 t + I$$

$$a = [NO_3^-]_0 - [Mo(V)]_0$$

$$k' \text{ shed} = k_2 K_2$$

Table I. Rate Constants for the Reduction of NO₃ by (NH₄)₂MoOCl₅

$10^{4}[(NH_{4})_{2}-M_{0}OCl_{5}]_{0}, M$	10^{2} [Et ₄ - NNO ₃] ₀ , M	10 ² [Na- NO ₃] ₀ , <i>M</i>	$10^{2} [NO_{3}]_{0}^{a} M$	10³[Et₄- NNC1], <i>M</i>	T, °C	$10^4 k_{obsd}^b/$ [NO ₄ ⁻] ₀ , sec ⁻¹	$10^{4} \cdot k'_{obsd}, c$	$ \frac{10^{4}k'' \operatorname{obsd}^{d}}{(K_{1} + [Cl^{-}])} \\ [Cl^{-}]/K_{1} \\ [NO_{3}^{-}], \operatorname{sec}^{-1} $
4.92		0.542	0.453		25	1.63	· · · · · · · · · · · · · · · · · · ·	
4.26		0.620	0.508		25	1.43		
5.07		1.57	1.07		25	1.48		
3.23		0.729	.581		25	1.45		
4.40		1.77	1.17		25	1.36		
5.69		3.10	1.76		25	1.41		
4.93		3.51	1.92		25	1.50		
5.38		5.00	2.43		25	1.70		
4.66		5.67	2.64		- 25	1.63		
3.59		6.67	2.91		25	1.50		
6.10		4.33	2.21		25	1.70		
2.61		3.03	1.73		25	1.74		
4.46		3.03	1.73		25	1.63		
5.23		3.03	1.73		25	1.95		
6.00		3.03	1.73		25	1.51		
7.53		3.03	1.73		25	1.60		
7.99		3.03	1.73		25	1.71		
9.99		3.03	1.73		25	1.54		
4.46		0.155	0.155		25		1.64	
5.94		0.104	0.104		25		1.74	
5.69	5.24				25	1.74		
4.61	5.18			0.490	25	1.82		
4.61	5.18			1.12	25	1.80		
5.23	5.18			2,12	25	1.76		
5.23	5.18			4.35	25	1.66		1.60
5.00	4.04			10.0	25			1.52
3.00	5.25			15.2	25			1.02
5.29	3.32			10.7	25			1.01
5.30	4.04			20.0	25			1.49
J.25 4.67	1.05			23.4	25	0 474		1.39
5.53	1.05				20	0.4/4		
3.33 4 97	1.11				30	2.01		
4.92	1.25				33	4.23		
4.15	1.20				50	· 0.3/		
7.15	1.05		• •		50	10.1		

^a $[NO_3^-]_0$ calculated from ion-pair equilibrium. ^b $k_{obsd} = k_3 K_2 [NO_3^-]_0$. ^c $k'_{obsd} = k_3 K_2$. ^d $k''_{obsd} = k_3 K_1 K_2 [NO_3^-]/[Cl^-](K_1 + [Cl^-]); \Delta H^{\ddagger} = 18.3 \pm 0.4 \text{ kcal mol}^{-1}; \Delta S^{\ddagger} = -14 \pm 2 \text{ cal mol}^{-1} \text{ K}^{-1} (25^\circ)$. $k_{obsd} / [NO_3^-]_0 = (1.63 \pm 0.15) \times 10^{-4} \text{ sec}^{-1} (25^\circ)$.

Plots of the data using these expressions are linear to >90% reaction and the values of the rate constants (k_3K_2) are consistent over the range of concentrations used (Table I).

When excess Cl⁻ is added, the rate expression becomes an ordinary psuedo-first-order expression, with $k''_{obsd} = k_3K_1K_2[NO_3^-]/[Cl^-](K_1 + [Cl^-])$. This may be rearranged to give

$$\frac{[NO_3^{-}]}{k''_{obsd}[Cl^{-}]} = \frac{[Cl^{-}]}{k_3K_1K_2} + \frac{1}{k_3K_2}$$

A plot of the left side of this equation vs. [Cl⁻] should be linear, with intercept $1/k_3K_2$ and slope $1/k_3K_1K_2$. This is in fact the case, as seen in Figure 1. Furthermore, the value of k_3K_2 obtained from the intercept ((1.56 ± 0.16) × 10^{-4} sec⁻¹) is in excellent agreement with the value obtained from runs without added Cl⁻ ($k = (1.63 \pm 0.15) \times 10^{-4}$ sec⁻¹).

The value of K_1 , evaluated from the slope of this plot is (3.6 \pm 0.8) \times 10⁻² M. This is in agreement with the assumption made in obtaining the integrated rate expression for runs in the absence of added Cl⁻ ($K_1 \gg$ [Cl⁻] since [Cl⁻] in those runs never exceeds $\sim 1.5 \times 10^{-3} M$. Furthermore, the value of K_1 indicates, at the concentration of (NH4)2MOCl₅ used, the dissociation in reaction 1 is essentially complete, in agreement with the assumption of [Cl⁻]₀ = [(NH4)2MOCl₅]₀ used in obtaining the integrated expression. The data, therefore, are consistent with the proposed mechanism over the range of concentrations used.

At high [Cl⁻], equilibrium 1 is displaced somewhat to the left, giving appreciable amounts of $MoOCl_2^{2-}$. The absorption

band and the molar absorptivity (ϵ) at 315 nm of solutions of (NH4)2MoOCl5 are insensitive to added Cl-, however, indicating MoOCl5²⁻ and MoOCl4⁻ are indistinguishable at this wavelength. This was verified by synthesizing NH4MoO-Cl4·CH3CN14 and comparing its spectrum in DMF with the spectrum of (NH4)2MoOCl5 in DMF. At the concentration used ($\sim 10^{-3}$ M) the two are essentially identical. This is perhaps not surprising, since the Cl- trans to oxygen in MoOCl5⁻² is known to be weakly bound, and it has been reported that its substitution by various ligands has little effect on the spectrum.¹⁸ Furthermore, recent work concerning MoOCl5²⁻ in nonaqueous solvents and in concentrated HCl indicates MoOCl4- is the major species present.¹⁹ Thus, the absorbance at 315 nm measures the total Mo(V) concentration, as required by the rate expression derived from the mechanism.

The activation parameters for the reaction, obtained from the Eyring plot of $\ln (k/T)$ vs. 1/T, are found in Table I. The rather large negative activation entropy (-14 cal) suggests considerable increase in order in the transition state. If the mechanism is correct, however, the experimental ΔS^{\dagger} involves ΔS for reaction 2 ($k_{exptl} = k_3 K_2$), which is unknown. Therefore, no conclusions concerning the origin of the entropy term may be drawn.

The biological reduction of nitrate may proceed by a mechanism similar to the one proposed here: a one-electron reduction of NO_3^- to NO_2 at a nonaqueous Mo(V) site followed by disproportionation of NO_2 to NO_3^- and NO_2^- in the aqueous environment after NO_2 is released by the enzyme. If the mechanism is correct, the rate constant for the actual

electron-transfer reaction, k_3 , must be considerably larger than k_{obsd} , since K_2 must be small. In the nitrate reductase enzymes, the preliminary displacement of a ligand by NO₃⁻⁻ might either be unnecessary or be much more favorable, accounting for the rapid enzymatic reduction rates.

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References and Notes

(1) R. C. Bray and J. C. Swann, Struct. Bonding, (Berlin), 11, 107 (1972). (2) R. H. Garrett and A. Nason, J. Biol. Chem., 244, 2870 (1969).

- (3) P. Forget and D. V. Der Vartanian, Biochim. Biophys. Acta, 256, 600 (1972).
- (4) P. Forget and D. V. Der Vartanian, Biochim. Biophys. Acta, 379, 74 (1975).
- (5) A. Nason Enzymes, 2nd Ed., 7, 587 (1963).
- J. T. Spence in "Metal Ions in Biological Systems", Vol. 5, H. Sigel, (6) Ed., Marcel Dekker, New York, N.Y., in press. V. Massey and H. Ganther, *Biochemistry*, **4**, 1161 (1965).
- J. T. Spence and E. P. Guymon, J. Phys. Chem., 70, 1964 (1966). (8)
- (9) J. T. Spence, Arch. Biochem. Biophys., 137, 287 (1970).
- (10) J. A. Frank and J. T. Spence, J. Phys. Chem., 68, 2131 (1964).
 (11) C. D. Garner, M. R. Hyde, F. E. Mabbs, and V. I. Routledge, Nature (London), 252, 579 (1974).
- (London), 252, 579 (1974).
 (12) C. D. Ritchie and G. H. Megerle, J. Am. Chem. Soc., 89, 1447 (1967).
 (13) W. G. Palmer, "Experimental Inorganic Chemistry", Cambridge University Press, Cambridge, England, 1934, p 408.
 (14) S. M. Horner and S. Y Tyree, Jr., Inorg. Chem., 2, 568 (1963).
 (15) M. R. Shinn, Ind. Eng. Chem., Anal. Ed., 13, 33 (1941).
 (16) P. Gray, R. Inst. Chem., Lect., Mongr., Rep., No. 4 (1958).
 (17) D. P. Ames and P. G. Sears, J. Phys. Chem., 59, 16 (1955).
 (18) E. A. Allen, B. J. Brisdon, D. A. Edwards, G. W. A. Fowles, and R. G. Williams, J. Chem. Soc. 4649 (1963).

- G. Williams, J. Chem. Soc., 4649 (1963).
- (19) P. M. Boorman, C. D. Garner, and F. E. Mabbs, submitted for publication.

Contribution from the Department of Chemistry, University of California, and the Inorganic Materials Research Division, Lawrence Berkeley Laboratory, Berkeley, California 94720

X-Ray Photoelectron Spectroscopic Study of Volatile Vanadium Compounds

RICHARD R. RIETZ, THEODORE F. SCHAAF, and WILLIAM L. JOLLY*

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Vapor-phase core electron binding energies were determined by X-ray photoelectron spectroscopy for nine vanadium compounds. Vanadium 2p3/2 binding energies were correlated using CHELEQ atomic charges and the "transition-state" point charge potential equation. Excellent correlation was obtained for compounds which are well represented by a single valence-bond structure. In the case of the other compounds, information regarding the relative contributions of various resonance structures and the importance of back-bonding was obtained by comparison of experimental and calculated binding energies.

Introduction

The CHELEQ electronegativity equalization procedure for calculating atomic charges has been successfully used, in conjunction with the point charge potential equation, to correlate core electron binding energies for a wide range of simple organic and inorganic compounds.^{1,2} The method has been parameterized for transition metal compounds and has been used to a limited extent for such compounds,^{2,3} but no systematic study has been made of the applicability of the method outside the main-group elements. In this paper we report gas-phase core electron binding energies determined by X-ray photoelectron spectroscopy for nine vanadium compounds and we discuss the nature of the bonding in these compounds as deduced from the data. This work is the first gas-phase XPS study of a series of compounds of a transition metal in which a wide range of metal oxidation states is spanned.

Experimental Section

Standard high-vacuum and inert-atmosphere techniques were employed. The organometallic vanadium compounds were handled under argon. All metal and glassware used in manipulation of the vanadium halides and oxyhalides were flamed under high vacuum and pretreated with the compound to be handled. Vanadium pentafluoride was prepared by the thermal disproportionation of VF4 (Orion) in a steel bomb.⁴ The VF₅ was twice treated with activated NaF⁵ and was stored in a bulb containing activated NaF. Vanadyl trifluoride (Alfa) was used without further purification. Vanadyl trichloride (ROC/RIC) was distilled through traps at 0, -78, and -196°. The clear liquid collected at -78° had a vapor pressure at 0° of 4.2 Torr (lit.⁶ 4.4 Torr). Vanadium tetrachloride (Orion) was distilled through traps at 0, -78, and -196°. The dark red liquid

collected at -78° was stored at -78° until used. Tris(hexafluoroacetylacetonato)vanadium,^{7,8} V(hfacac)₃, was prepared by the method of Morris et al.8 and purified by sublimation (mp 75.5-76.5°, lit.7 76°). The $(\pi$ -C₅H₅)V(CO)4 (Strem) (mp 139.5–140.5°, lit.⁹ 139°) was used without further purification. Its infrared spectrum agreed with the literature.9 Vanadium hexacarbonyl was prepared from $[Na(diglyme)_2][V(CO)_6]$ (Orion) by the method of King.¹⁰ It was stored in the dark at -78° over PsOs and resublimed immediately before use. The $(\pi$ -C₇H₇)V(CO)₃ was prepared from V(CO)₆ and cycloheptatriene by the method of Werner and Manastyrskyj11 and was purified by sublimation. The product melted at 133-134° (lit.11 134-137°) and exhibited an infrared spectrum in agreement with the literature.¹¹ The $(\pi$ -C₇H₇)V(π -C₅H₅) was prepared from (π - $C_5H_5)V(CO)_4$ and cycloheptatriene by the method of King and Stone¹² and was purified by sublimation at 120°.

X-Ray Photoelectron Spectra. Spectra were obtained using the Berkeley iron-free, double-focusing magnetic spectrometer.¹³ The halides, oxyhalides, and vanadium hexacarbonyl were irradiated in a gold-plated chamber with a volume of 300 cm³, a 0.5-mm exit slit, and an aluminum X-ray window. Tris(hexafluoroacetylacetonato)vanadium, $(\pi$ -C₅H₅)V(CO)₄, and $(\pi$ -C₇H₇)V(CO)₃ were irradiated using a slit plug described by Banna.¹⁴ The sample and reference gas were handled in a system constructed of 0.5-in. stainless steel pipe. This system was assembled and secured while back-flushing with argon gas as in standard Schlenk manipulation. The vapor of the compound passed to the irradiation chamber through a 15-in. length of tubing constructed of Pyrex glass and stainless steel. The samples of VCl4 and V(CO)6 were held at 0° during data collection to minimize decomposition. The $(\pi$ -C₇H₇)V $(\pi$ -C₅H₅) was held in a heated chamber¹⁴ at ca. 100°.

Magnesium K α X-rays (1253.6 eV) were used for all spectra. Argon or neon at a partial pressure of $20-40 \ \mu$ was introduced with the sample to the irradiation chamber, and the Ar $2p_{3/2}$ line (E_B = 248.45 eV) or the Ne 1s line ($E_B = 870.21 \text{ eV}$) was used as a reference.